

Setting the Standard in rAAV Manufacturing: A Comparative Bioreactor Study Showcases the Strength of the EpyQ® STR Platform

M Boscher, J Wagner, J Babic, T Fischer, B L Carnio, C Zach, B Hager, M Gora, M Hoerer, K Heller, A Youssef

Ascend Advanced Therapies GmbH, Process Development, Munich, Germany

Introduction

Recombinant Adeno-Associated Virus (rAAV) vectors are shaping the future of gene therapy. Lowering production costs and boosting efficiency remain key to enabling broader access and driving market growth.

Using our proprietary split 2 plasmid system (EpyQ®), we consistently achieve $>1 \times 10^{11}$ vg/mL across multiple rAAV serotypes and scales (250 mL – 200 L) in stirred-tank bioreactors, demonstrating reliable, high-yield production. Although stirred tanks deliver strong performance in our hands, benchmarking against alternative bioreactor systems ensures we continue using the optimal platform. In our experience, orbital shaking and rocking

bioreactors support robust HEK293 growth while applying lower power input to the cells, making them promising platforms for efficient rAAV manufacturing.

In this study, we compared rAAV9 production across three platforms - orbital shaker (SB10-X single-use Bioreactor; Kuhner), rocking bioreactor (ReadyToProcess™ WAVE 25 Rocker; Cytiva), and stirred-tank (Biostat® B 10 L single-use Bioreactor; Sartorius) - to evaluate how bioreactor design and operating conditions (shear, mixing, power input) affect cell growth, vector yield, and product quality.

rAAV Production & Quality Workflow

Upstream

- Platforms: orbital shaking, rocking, and stirred-tank bioreactors
- Serotype: rAAV9
- Scale: 10 L (equal working volumes)
- Cell line: proprietary HEK293
- Transfection: EpyQ split 2 plasmid system

Offline Monitoring

- Cell density & viability (ViCELL-BLU)
- Metabolic profiles (BioProfile® Flex2)

Analytics

- Vector yields in crude lysates: vg titer (ddPCR), capsid yields (Gyrolab® immunoassay)
- % Full capsids (mass photometry, Refeyn)
- Encapsidated plasmid- and host-cell DNA-derived impurities (ddPCR)
- In-vitro vector potency relative to reference

Orbital Shaking



SB10-X single-use Bioreactor

Rocking Motion



ReadyToProcess™ WAVE 25 Rocker

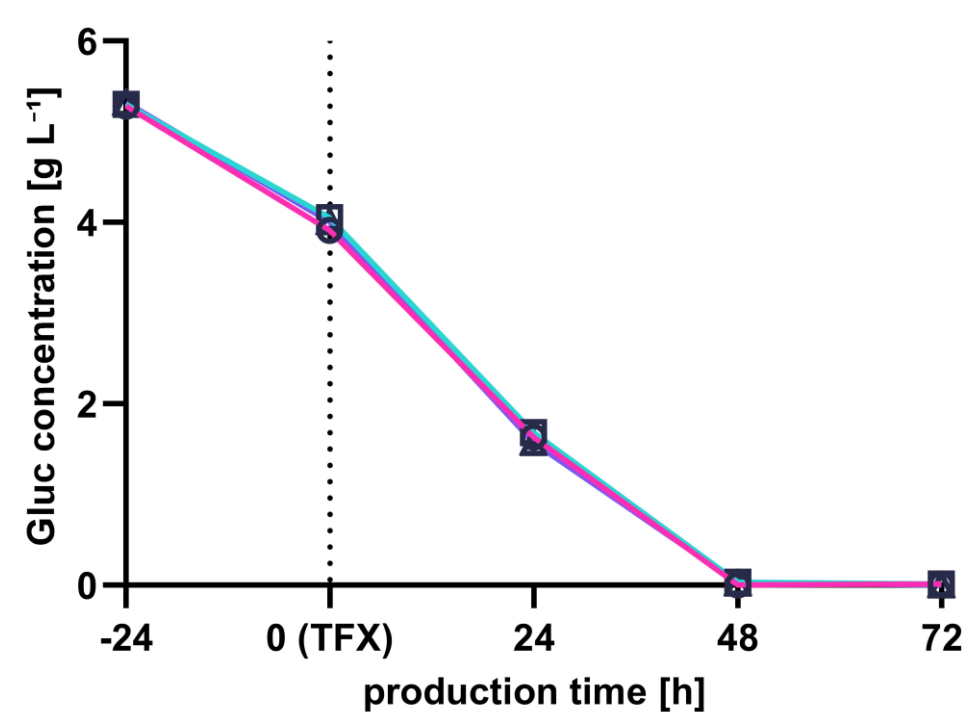
Stirring



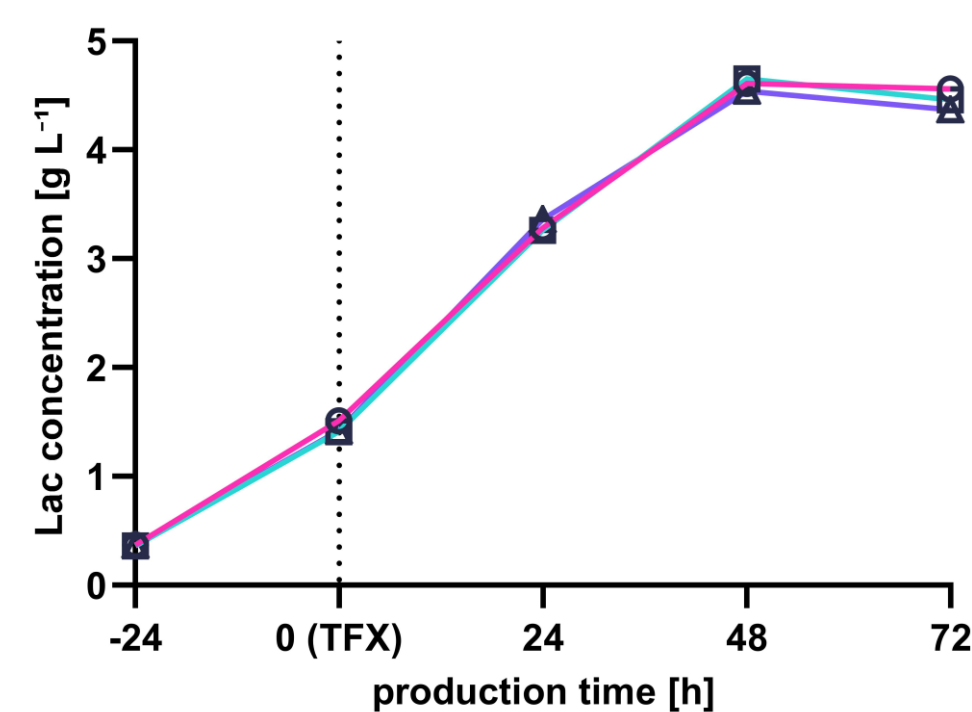
Biostat® B 10L single-use Bioreactor

Metabolite Trends

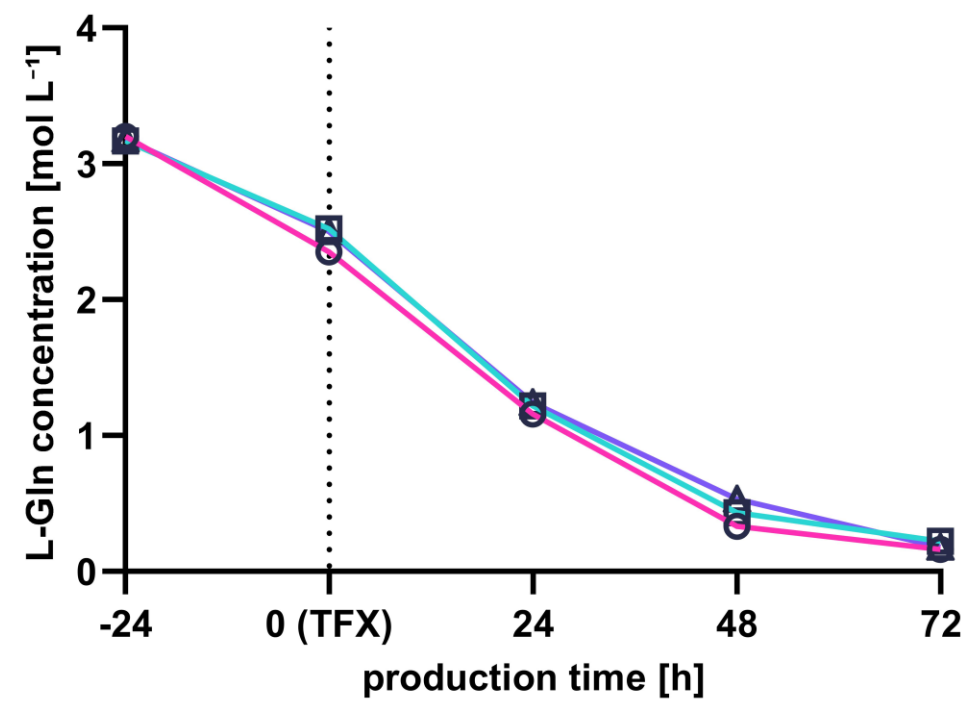
Glucose Consumption



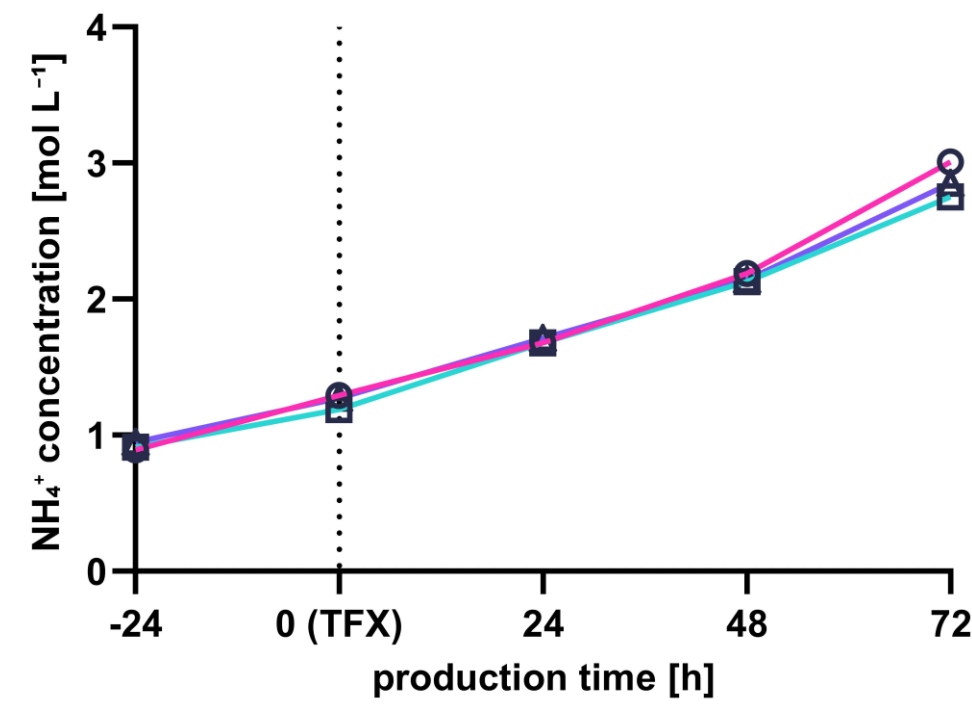
Lactate Accumulation



Glutamine Consumption



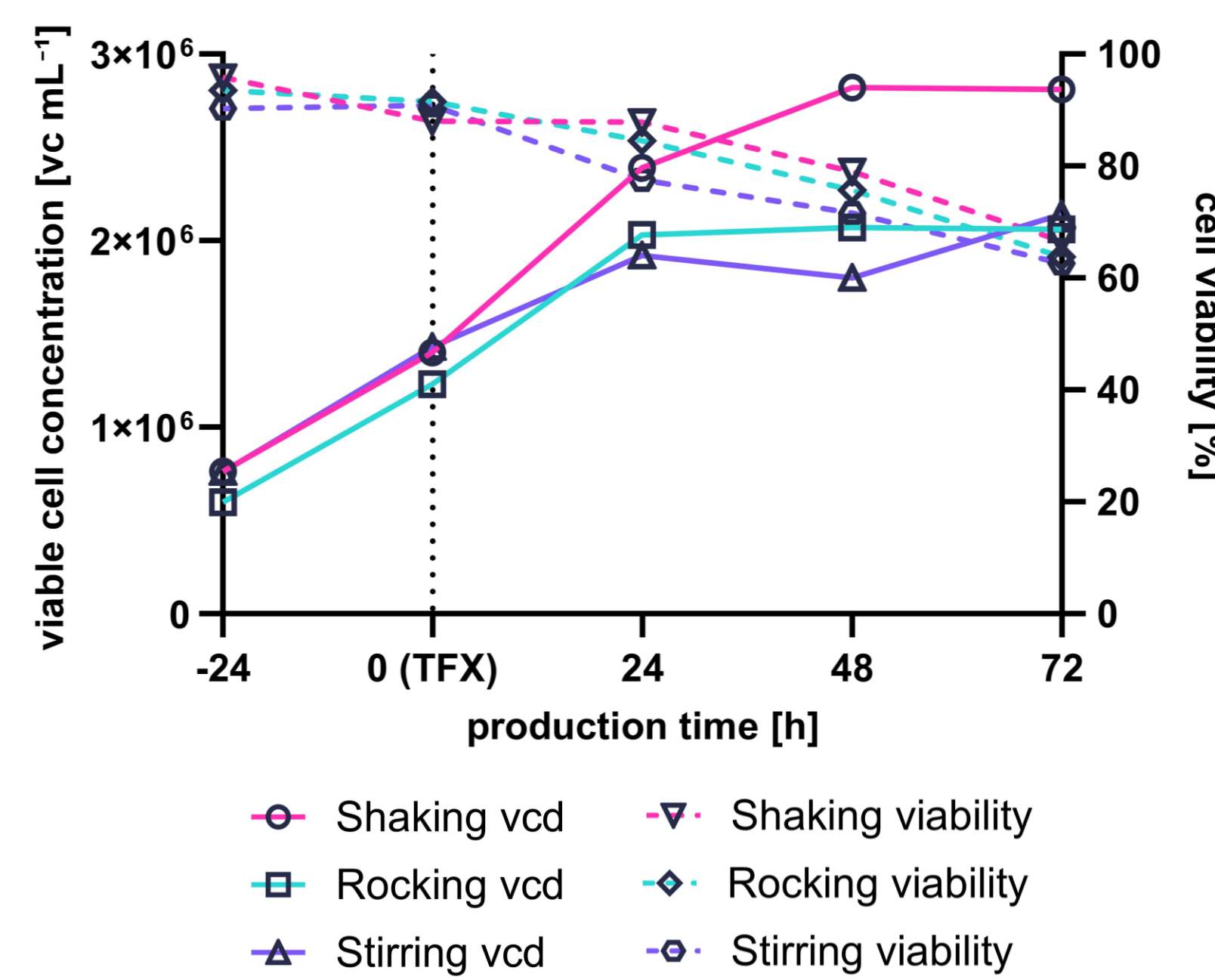
NH₄⁺ Accumulation



○ Shaking □ Rocking ▲ Stirring

Growth and viability

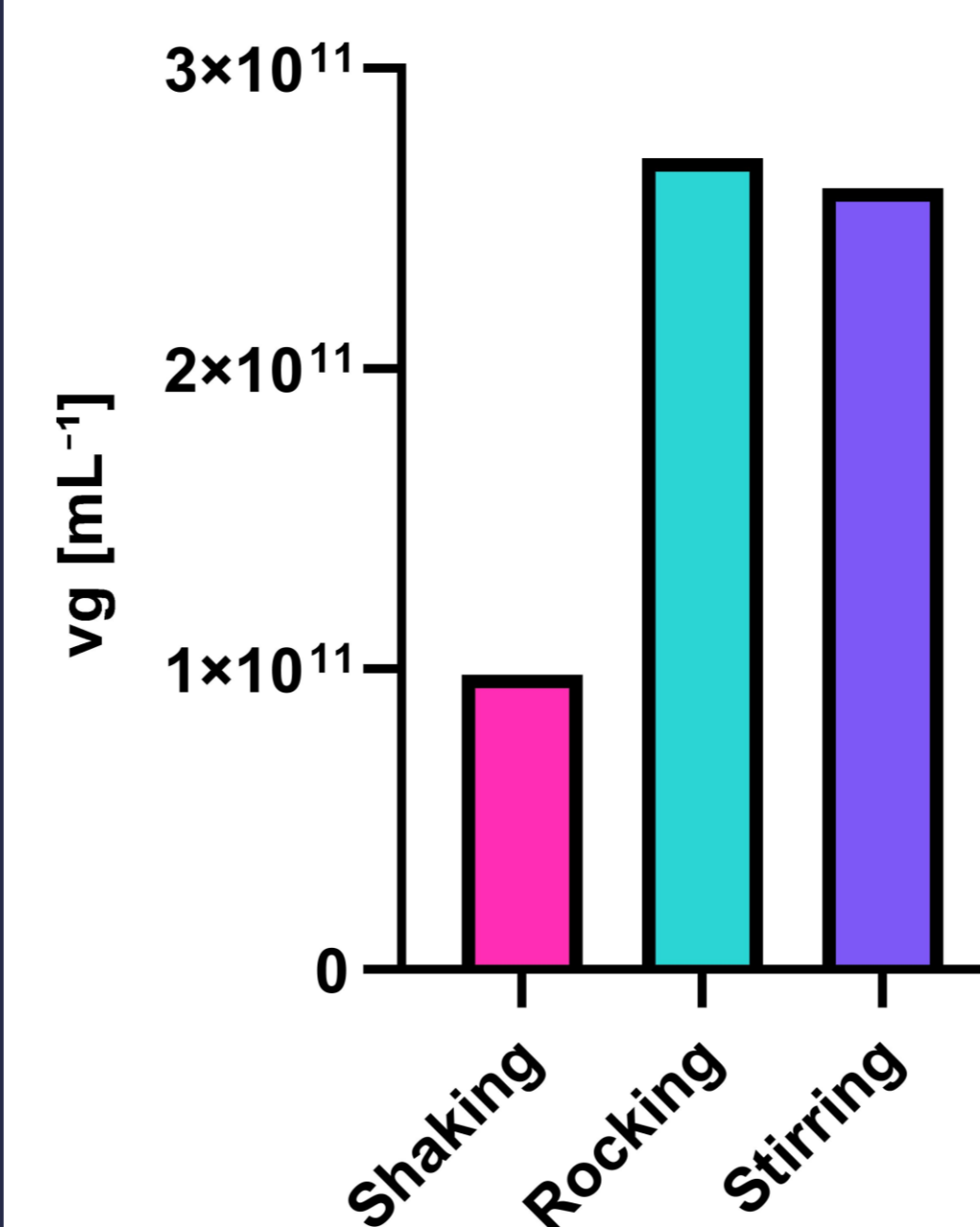
Viable Cell Density and Viability



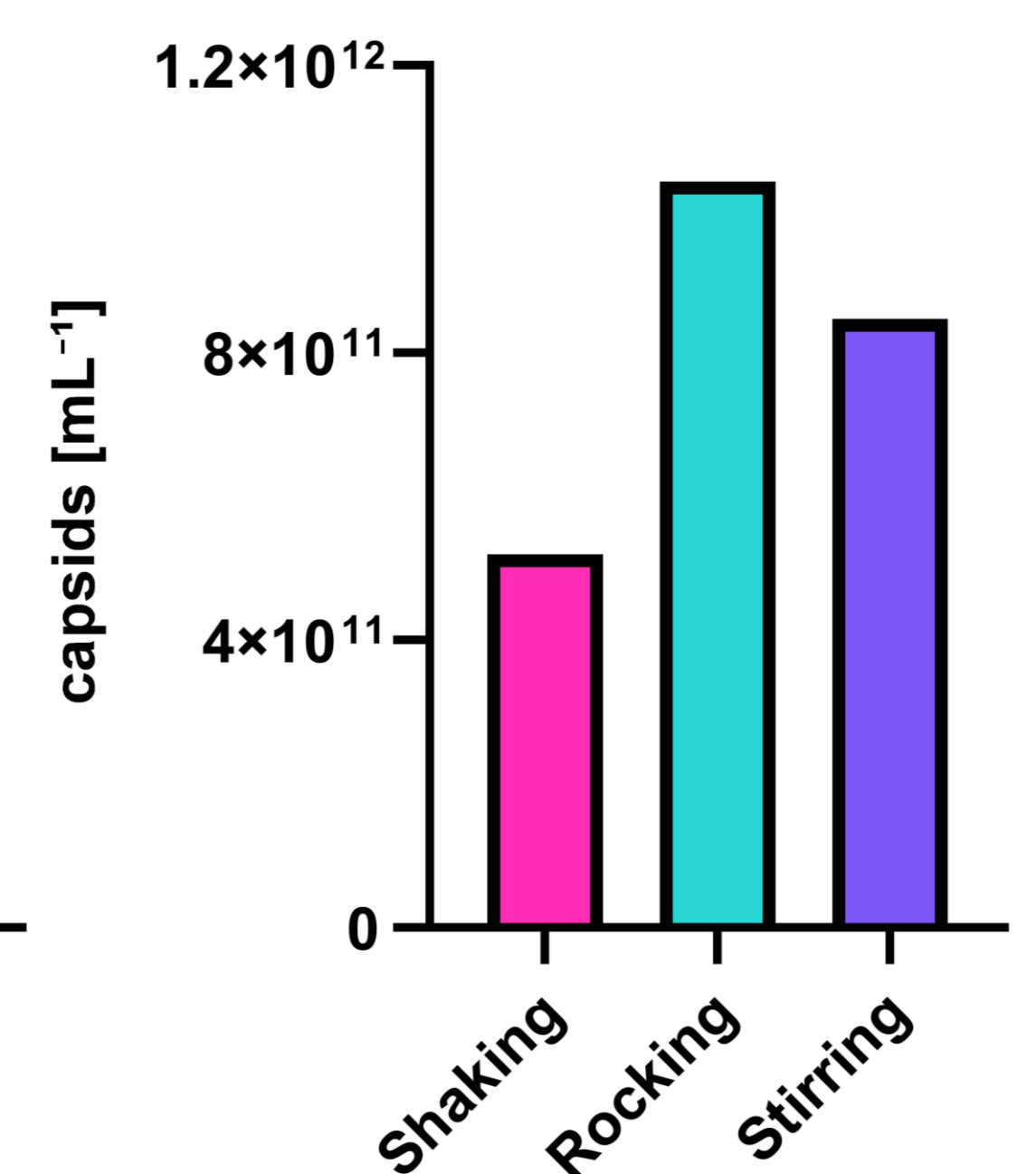
○ Shaking vcd □ Rocking vcd ▲ Stirring vcd
● Shaking viability ◆ Rocking viability ▲ Stirring viability

Crude Lysate Titers

Vector Genome Yields

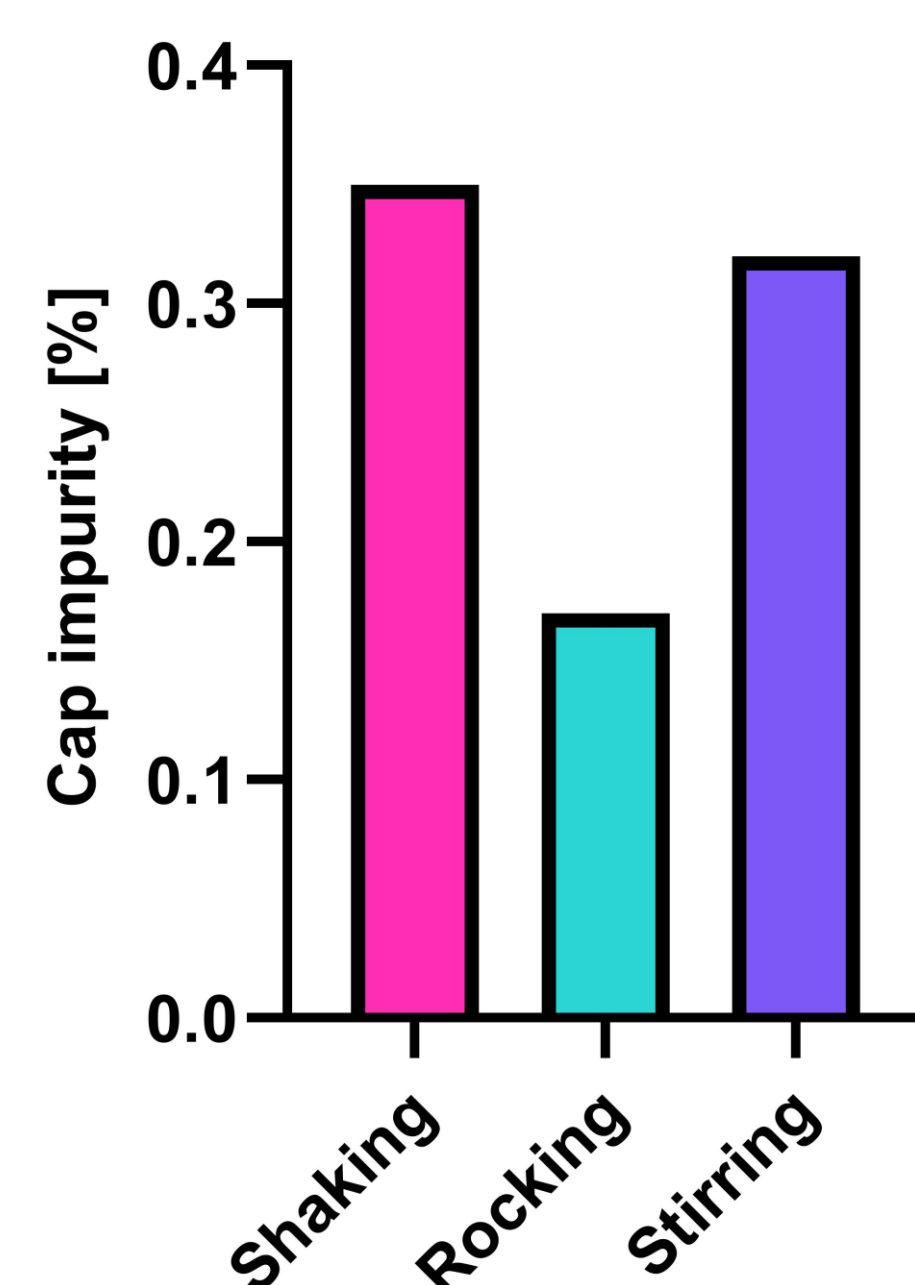


Capsid Yields

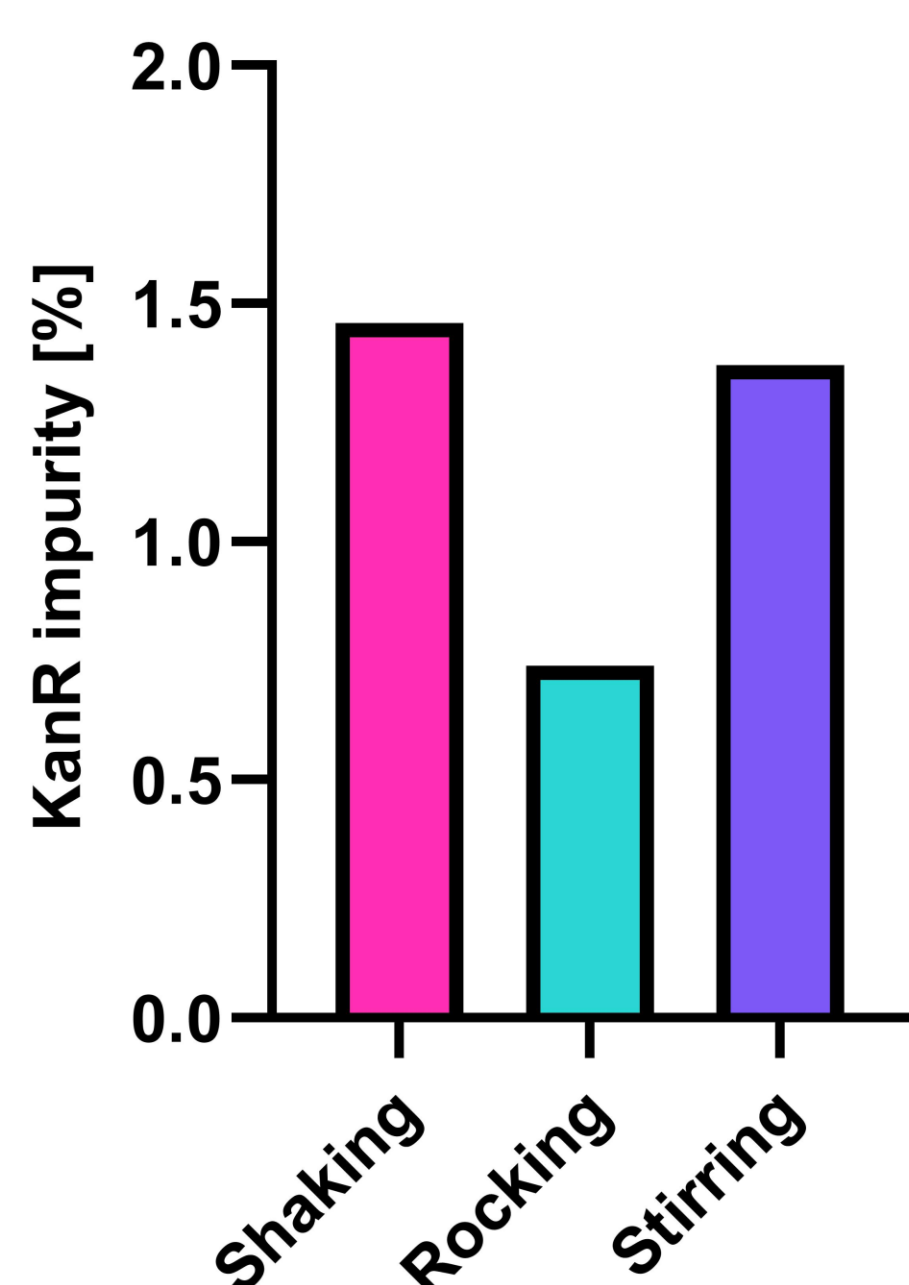


Encapsidated Impurity Profiles

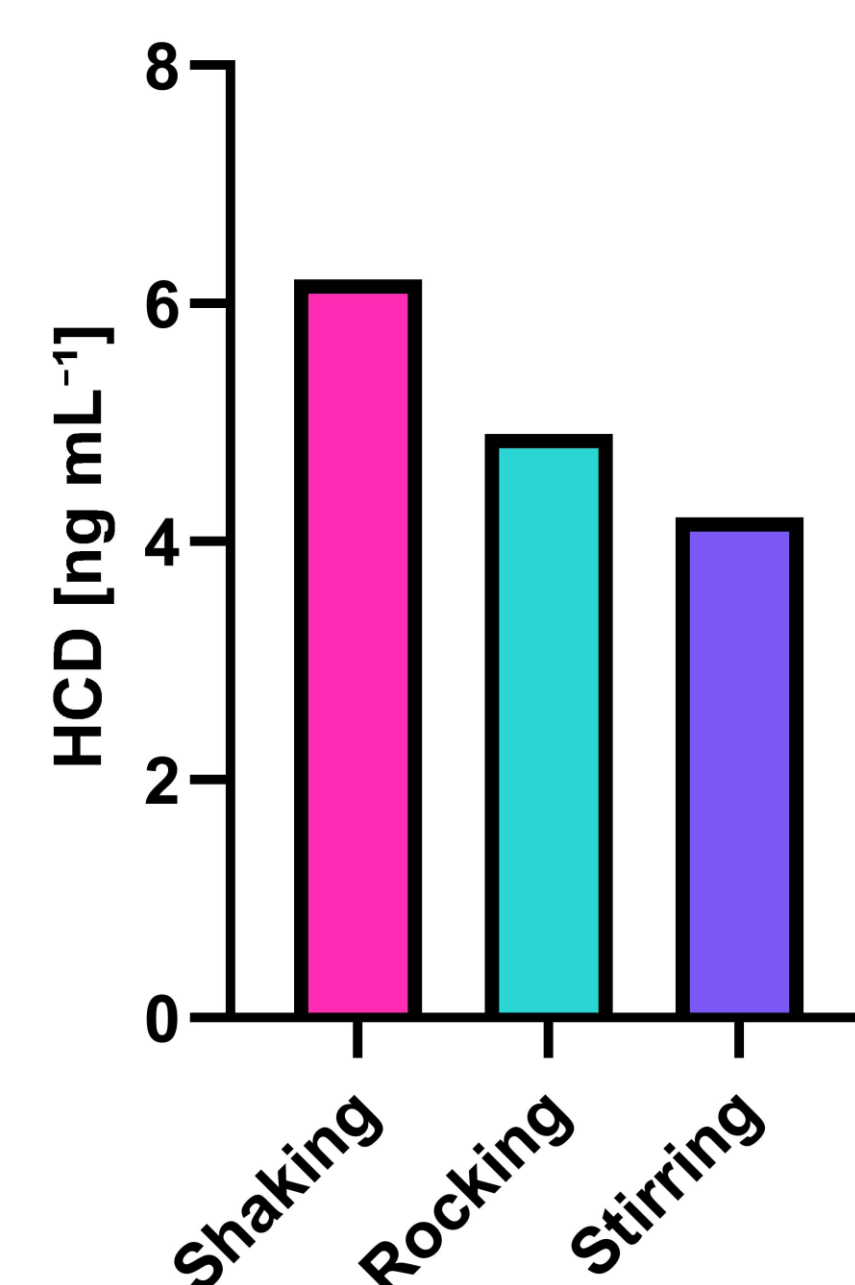
Capsid Gene-Derived Impurities



Kanamycin Resistance Gene-Derived Impurities

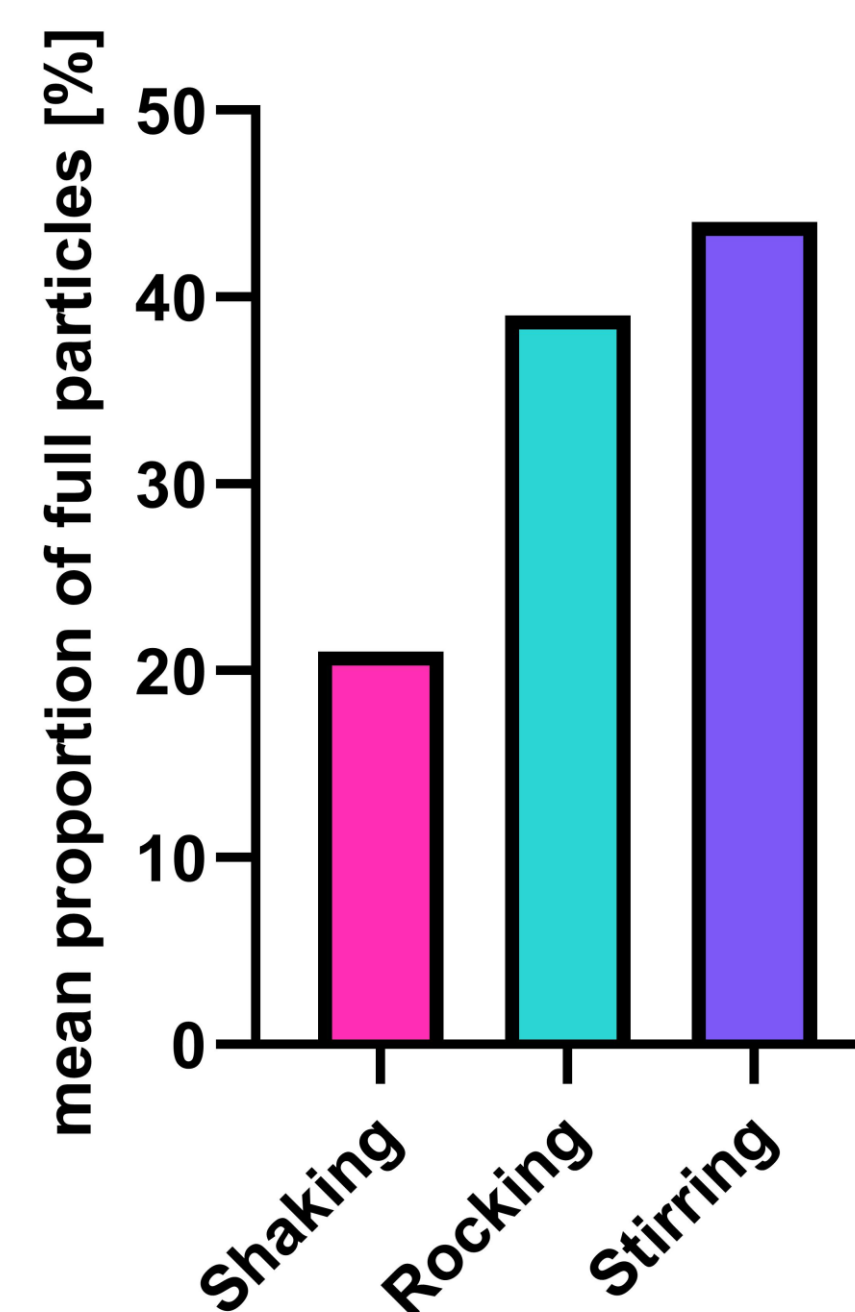


Host-Cell DNA-Derived Impurities



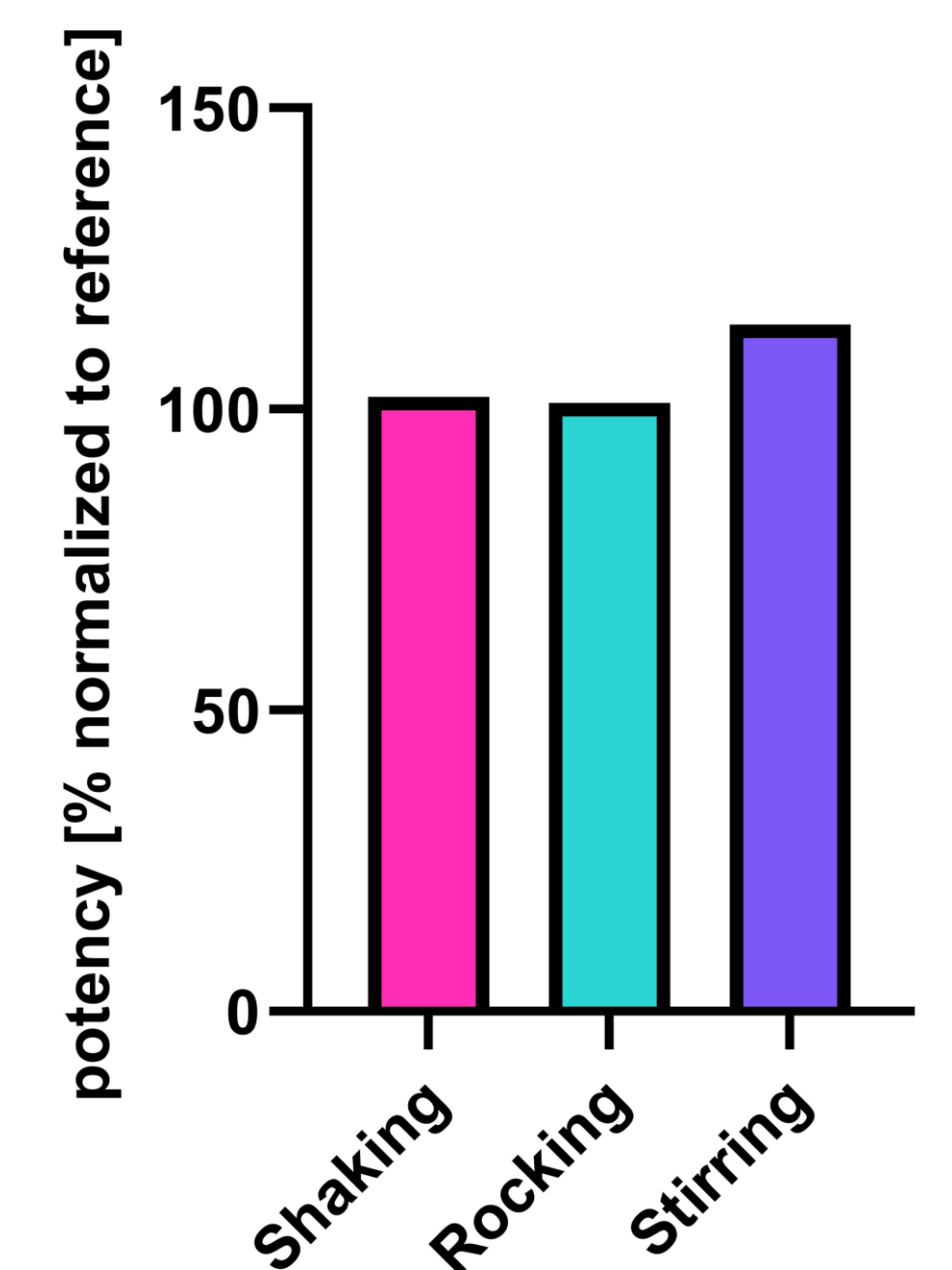
Full Capsid Yield

Full/Empty Proportion



Biological Performance of the Drug Substance

In Vitro Potency



Summary

Both stirred-tank and rocking bioreactors demonstrated excellent performance in rAAV production, clearly outperforming the shaken system in yield and product quality.

In-process data were largely consistent across platforms, with only the shaking system showing prolonged post-transfection growth, possibly reducing productivity.

While the rocking bioreactor showed strong results, its limited scalability confirms the stirred-tank bioreactor as our preferred platform for robust, large-scale manufacturing using our proprietary HEK293 cell line with the EpyQ platform.

